Biocompatible fluorescent silicon nanocrystals for single-molecule tracking and fluorescence imaging

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Fluorescence microscopy is used extensively in cell-biological and biomedical research, but it is often plagued by three major problems with the presently available fluorescent probes: photobleaching, blinking, and large size. We have addressed these problems, with special attention to single-molecule imaging, by developing biocompatible, red-emitting silicon nanocrystals (SiNCs) with a 4.1-nm hydrodynamic diameter. Methods for producing SiNCs by simple chemical etching, for hydrophilically coating them, and for conjugating them to biomolecules precisely at a 1:1 ratio have been developed. Single SiNCs neither blinked nor photobleached during a 300-min overall period observed at video rate. Single receptor molecules in the plasma membrane of living cells (using transferrin receptor) were imaged for ≥10 times longer than with other probes, making it possible for the first time to observe the internalization process of receptor molecules at the single-molecule level. Spatial variations of molecular diffusivity in the scale of 1–2 µm, i.e., a higher level of domain mosaicism in the plasma membrane, were revealed.

Introduction

Fluorescence microscopy is one of the most important tools for cell-biological and biomedical research as well as clinical diagnostics (Torres et al., 2008; Toomre and Bewersdorf, 2010; Yan et al., 2011; Germain et al., 2012). However, three major problems with the presently available fluorescent probes still often appear in fluorescence microscopy: (1) photobleaching, (2) blinking (Dickson et al., 1997; Yao et al., 2005), which almost all of the presently available probes exhibit, and (3) the large size of GFP and quantum dots (QDs). Extensive efforts have been made to produce QDs and organic fluorescent molecules to alleviate these problems (Buschmann et al., 2003; Howarth et al., 2008; Muro et al., 2010; Altman et al., 2012a,b; Liu et al., 2012), but their applicability to living cells, particularly for single-molecule imaging using living cells (Cognet et al., 2006; Smith and Nie, 2010; Kasai et al., 2011), has been severely limited because the problems of photobleaching, blinking, and cross-linking target molecules are more critical in single-molecule imaging in living cells (Ha and Timefeld, 2012).

Physicists and chemists have known for almost two decades that silicon nanocrystals (SiNCs) as small as 1–5 nm in diameter are fluorescent (Canham, 1990; Yamani et al., 1998; Credo et al., 1999; Akcakir et al., 2000), with an emission spectrum that varies with the particle size, from blue from the smallest to red from the largest (Belomoin et al., 2002; Zhou et al., 2003; Syuchugov et al., 2005a), do not easily photobleach (Gelloz et al., 2003; Warner et al., 2005), and could be made water soluble (Li et al., 2004; Li and Ruckenstein, 2004; Warner et al., 2005) and targeted to biological molecules (Wang et al., 2004; Erogboho et al., 2008, 2011a,b). Recently, a possibility of greatly improving the quantum efficiency of SiNCs has been proposed (de Boer et al., 2010).
Results

Preparing mercaptosilane-coated SiNCs with an ~4-nm hydrodynamic diameter

Inspired by the work of Yamani et al. (1998), Akcakir et al. (2000), and Belomoin et al. (2002), we developed an easier protocol that specifically generates red-emitting SiNCs (see Materials and methods subsections Preparation of SiNC on the wafer through Recovery and purification of mSiNC; Fig. 1 A–C). This method employs the simple chemical etching of silicon wafers (Fig. 1 A), rather than the electrochemical etching used previously. Then, the SiNCs, still attached to the wafer (wSiNCs), were coated by conjugating 3-mercaptopropylthiolsilane (3-MPTS) to the hydroxyl (silanol) groups on the wSiNC surface (wmSiNCs; Fig. 1 B). Coating was confirmed by Fourier transform infrared (FTIR) spectroscopy (Fig. 2 A, top). The fluorescence spectrum of wSiNC and wmSiNC are shown in Fig. 2 B. The fluorescence spectrum changed slightly after hydrophilic coating. The SiNCs were then liberated from the polished side of the wafer by mechanical scraping into water. This suspension was centrifuged briefly to remove the large debris and then was purified by gel filtration HPLC (mSiNCs; Fig. 1 C). FTIR spectroscopy confirmed that the 3-mercaptopropylthiolsilane (3-MPTS) remained on the mSiNC surface after this procedure (Fig. 2 A, bottom).

In the gel filtration HPLC using a Superdex 75 column, the purified mSiNCs eluted in a sharp band between cobalamin...
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be 4.1 ± 0.14 nm with a polydispersity index of 0.14 ± 0.028 (mean ± SD throughout this paper, where SD represents the SD of the mean value for \( n \) independent experiments; \( n = 3 \) here), indicating a sharp distribution of the particle size (Fig. 3 C, curve). It is concluded that, after Superdex 75 gel filtration HPLC, the particle size was uniform, and the contamination of larger particles, mSiNC dimers, and greater clusters was very limited.

The mSiNC in aqueous buffer was further characterized by fluorescence correlation spectroscopy (FCS; see FCS and determination of the mSiNC size based on the FCS measurements in Materials and methods). Typical plots of the autocorrelation function versus time for mSiNC, GFP, Qdot655, and rhodamine 6G are shown in Fig. 3 D. The hydrodynamic diameter of mSiNC was determined to be 4.1 ± 0.14 nm with a polydispersity index of 0.14 ± 0.028 (mean ± SD throughout this paper, where SD represents the SD of the mean value for \( n \) independent experiments; \( n = 3 \) here), indicating a sharp distribution of the particle size (Fig. 3 C, curve). It is concluded that, after Superdex 75 gel filtration HPLC, the particle size was uniform, and the contamination of larger particles, mSiNC dimers, and greater clusters was very limited.

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Transmission EM (TEM) images of the mSiNCs recovered from the gel filtration peak (Fig. 3 E) provided the mSiNC diameter distribution (Fig. 3 F), which can be fitted by a Gaussian function (3.2 ± 0.4 nm; n = 278 particles). Because this diameter represents that of the mSiNC core without including the surface coating, and in addition, the hydrodynamic diameter represents...
that of the hydrodynamic slipping layer around mSiNC, the smaller TEM diameter is consistent with the 4.1-nm hydrodynamic diameter determined by DLS and FCS.

A fluorescence image of a cuvette containing mSiNCs purified by gel filtration HPLC, excited with a UV lamp, is shown in Fig. 2 C. The absorption and fluorescence spectra of the purified mSiNCs are shown in Fig. 2 (D and E). The peak of the fluorescence spectrum is ~655 nm, and its width is broader (full width at half maximum = ~100 nm) than that for the QDs by a factor of 2–3, consistent with a previous finding that even a uniform population of blue-emitting SiNCs of 1.4 ± 0.3 nm in diameter exhibited the fluorescence spectrum with a full width at half maximum of ~80 nm (Warner et al., 2005).

Storing the mSiNCs in dehydrated benzene at 4°C for 60 d did not affect either their fluorescence spectrum or fluorescence intensity (Fig. S1), as also reported for hydrophobic SiNC in toluene (Li et al., 2004). However, in aqueous solutions, the fluorescence intensity was reduced much faster. The fluorescence intensity of mSiNC conjugated with a single molecule of transferrin (Tf; mSiNC-Tf) decreases with an exponential decay time of 42 ± 2.8 h in PBS at 37°C (n = 4; the fitting error at the 68.3% confidence limit is given; see Long-term storage mSiNC and the preparation and observation of mSiNC-Tf in aqueous solutions during a day in Materials and methods; Fig. S2).

Each individual mSiNC neither blinks nor photobleaches

To determine whether each mSiNC blinks or photobleaches, purified mSiNCs were adsorbed and immobilized on polystyrene-coated coverslips and were imaged with a home-built objective lens–type total internal reflection fluorescence (TIRF) microscope (a 440-nm laser line with 0.34 µW/µm² on the sample plane; Fig. 4 A; Koyama-Honda et al., 2005). The distribution of the fluorescence intensities of individual mSiNCs (log-normal fitting; Mutch et al., 2007; Fig. 4 B, red line) is consistent with the presence of a population of single fluorescent particles (Fig. 4 B, top). Meanwhile, the fluorescence intensities of single individual Qdot655 particles exhibited much greater variations (Fig. 4 B, bottom).

A representative plot of the fluorescence intensity versus time of a single mSiNC at a 33-ms (video rate) resolution is shown in Fig. 4 C (top) and should be compared with that of a CdSe-QD (Qdot655 here) observed under exactly the same illumination and detection conditions (Fig. 4 C, bottom). Qdot655 exhibited frequent blinking (for the nature of the blinking and its mechanism for QDs, see Chen et al. [2008], Mahler et al. [2008], and Wang et al. [2009]) as well as gradual photobleaching (Fig. 4 C and Fig. S3 B), consistent with previous observations (van Sark et al., 2002). None of the 30 mSiNCs observed under the same conditions blinked or photobleached (during the total period of 300 min), although Sychugov et al. (2005b) reported the blinking of SiNCs. Under the conditions in which similar signal-to-noise ratios are attained for single fluorescent spots, single molecules of GFP or the organic dye Cy3 photobleached within ~3 s (unpublished data). Namely, by using mSiNCs as fluorescent probes, the observation time of single molecules could be lengthened by a factor of 6,000 or more (without a single blinking event), eliminating the two major roadblocks for single-molecule tracking.

The average fluorescence signal intensity for a single mSiNC particle, as determined by the log-normal fitting of the distribution of signal intensities of single mSiNCs (Fig. S4 A) is increased linearly at least ≤0.68 µW/µm² on the sample plane (Fig. S4 B), twice the standard laser power used in this work. No photobleaching and blinking were observed even at 0.68 µW/µm².

We noted that many QDs suddenly started emitting fluorescence sometime after irradiation with the excitation light, consistent with a previous study (for example, see Li-Shishido et al., 2006). This result suggests that significant fractions of Qdot655 and mSiNCs might be nonfluorescent at a given time. In fact, Credo et al. (1999) reported that only 2.8% of SiNCs are fluorescent.

Therefore, we examined the fractions of nonfluorescent mSiNCs using Alexa Fluor 488–conjugated mSiNC (see Preparation of Alexa Fluor 488–labeled mSiNC [Alexa Fluor 488–mSiNC] and determination of the fraction of mSiNCs that are actually fluorescent in Materials and methods; Fig. 5). The result showed that 38.4 ± 2.6% of the mSiNCs were actually fluorescent (for six independently prepared specimens, examining 245 Alexa Fluor 488 spots in total), indicating much improvement from the previously reported value of 2.8%, although the improvement mechanism is not clear.

Extinction coefficient and quantum yield of mSiNCs

FCS (Fig. 3 D) can determine the concentration of mSiNC particles that are actually fluorescent. Meanwhile, the result described in the previous subsection indicates that 38% of mSiNC particles fluoresce after they absorb light (making a plausible assumption that nonfluorescent particles have the same extinction coefficient as fluorescent ones). Using the absorption spectrum (Fig. 2 D, black solid line) and the concentration of mSiNCs (the concentration determined by FCS divided by 0.38), the extinction coefficients were obtained as 10.2 (±2.3) × 10², 5.7 (±1.1) × 10², and 5.3 (±1.4) × 10² M⁻¹ cm⁻¹ at 350, 400, and 450 nm, respectively (n = 12). This is the first determination of the SiNC extinction coefficients in an aqueous solution, and it shows that the mSiNC’s extinction coefficient is comparable to those of protein fluorophores, although these values are substantially smaller than those for QDs (Table S1).

As another method to determine the extinction coefficient, we evaluated the molar concentration of mSiNC in aqueous solutions without relying on its optical properties (such as FCS and the fraction of actually fluorescent particles), we measured the actual weight of mSiNC after drying and estimated the molecular weight of the 3.2-nm mSiNC (36,083.6; see Determination of the amount of mercaptosilane bound to, and the extinction coefficient of, mSiNC in Materials and methods; in this process, the mean number of 3-MPTS molecules coating the mSiNC surface was determined as 51.2 ± 19.4 molecules/mSiNC particle [n = 3]). From the absorption spectrum obtained for these solutions, the extinction coefficients at 350, 400, and 450 nm of the mSiNC solution were estimated to be 10.5 (±2.2) × 10², 4.6 (±1.5) × 10², and 3.7 (±1.3) × 10² M⁻¹ cm⁻¹ (n = 3),
The fluorescence quantum yield of the mSiNCs in 50 mM phosphate buffer, pH 7.0, was determined to be 0.080 ± 0.0060 (n = 6) (see Determining spectroscopic properties of mSiNC). Because this value was determined for the bulk mSiNC suspension (and thus is listed as bulk in Table S1), it includes mSiNCs that do not fluoresce even after absorbing light (~62%). The quantum yield of fluorescent mSiNCs was obtained by multiplication by 1/0.384, giving a quantum yield of fluorescent mSiNCs of 0.21 ± 0.016 (n = 6). When each individual mSiNC particle is to be observed, the quantum yield of 0.21 should be used, and therefore, it is listed as single particle in Table S1. As shown in Table S1, the quantum yield of mSiNC (0.21) is smaller than those for Qdot655 (>0.50) but is comparable with representative fluorescent organic Cy dye molecules. It is greater than the quantum yield of Cy3 (0.04) but smaller than that of Cy5 (0.27; measured in PBS; Mujumdar et al., 1993).
extinction coefficients from the supplier could not be verified in any refereed publications). However, as shown in Fig. 4 C and Fig. S3, initially, the Qdot655 is brighter, but because it blinks frequently and its fluorescence intensity varies greatly with time, whereas the mSiNC fluorescence is stable over the same period, the fluorescence intensities of Qdot655 and mSiNC, averaged over the entire duration in which the Qdot655 emits a fluorescence signal (Fig. 4 C and Fig. S3, red, solid, horizontal lines), were similar to each other, under exactly the same illumination and detection conditions. Because single-molecule imaging has been widely conducted using, for example, EGFP (quantum yield $\phi$ of $\sim$0.6 and molar extinction coefficient $\varepsilon$ of $\sim$55,000 M$^{-1}$ cm$^{-1}$; McRae et al., 2005) and Cy3 ($\phi$ of $\sim$0.04 and $\varepsilon$ of $\sim$150,000 M$^{-1}$ cm$^{-1}$; Table S1), it is not surprising that mSiNC ($\phi$ of $\sim$0.21 and $\varepsilon$ of $\sim$55,000) can be observed at the level of single particles. This suggests that the extinction coefficient of Qdot655 provided by the supplier must be cautiously reevaluated.

### Conjugating mSiNCs to proteins

Using the protocol shown in Fig. 6 (A and B), mSiNC was conjugated to Tf, a serum protein that carries ferric ions into cells (see Preparation of mSiNC-labeled Tf [mSiNC-Tf] in Materials and methods). To examine the progress of the chemical conjugation, the reaction mixture of maleimide-conjugated Tf (maleimide-Tf) and mSiNC (2:1 mol ratio; 0 and 60 min) was subjected to Superdex 200 gel filtration HPLC chromatography, and its elution pattern is shown in Fig. 6 C (blue dextran represents the void volume). The mSiNC peak (Fig. 6 C, peak 3) decreases as the conjugation proceeds, whereas peak 2 grows and shifts toward larger molecular size. The behavior of the peak 2 (Fig. 6 C) suggests that the peak 2 after reaction for 60 min probably represents partially overlapped elution of Tf conjugated with mSiNC (mSiNC-Tf; larger) and maleimide-Tf (smaller). Peak 1 (Fig. 6 C) did not change. Because we suspected the presence of maleimide-Tf dimers in the reaction mixture, we generated artificial Tf dimers by a chemical cross-linker, bis(sulfosuccin imidyl)suberate, from Tf monomers (Tf monomers and dimers were confirmed by SDS-PAGE) and examined the Superdex 200 chromatogram of the reaction mixture (Fig. 6 C, bottom). From this elution pattern, we concluded that peak 1 in Fig. 6 C (top) represents maleimide-Tf dimers. Because the shape and the position of peak 1 did not change appreciably after the mSiNC conjugation reaction for 60 min, it is concluded that mSiNC does not induce cross-linking of maleimide-Tf under these conjugation conditions.

Then, we tried to separate mSiNC-Tf and maleimide-Tf (overlapped peak 2 on the Superdex 200 chromatogram shown in Fig. 6 C, top) using Superdex 75. The Superdex 75 gel filtration HPLC chromatograms of the reaction mixture of maleimide-conjugated Tf and mSiNC (2:1 mol ratio; 0, 30, and 60 min), shown in Fig. 6 D, reveal the progress of the chemical conjugation. The maleimide-Tf and mSiNC peaks (Fig. 6 D, peaks 2 and 3, respectively) decrease as the conjugation proceeds, whereas the new peak 1 (Fig. 6 D) at a larger molecular size appears and grows (the small peak eluted slightly after peak 1 at time 0 probably represents small amounts of contamination in the reaction mixture), suggesting that this new peak represents mSiNC-Tf.

Next, we examined whether peak 1 in the Superdex 75 chromatogram (Fig. 6 D) contained Tf molecules bound by multiple mSiNCs. To investigate this possibility, we determined the number of mSiNCs attached to each Tf molecule. First, the mSiNC-Tf complexes recovered in peak 1 (Fig. 6 D) were adsorbed and immobilized on coverslips, and then, they were observed at the single-particle level using TIRF microscopy. The distribution of the signal intensity of each mSiNC-Tf spot after 30- and 60-min reactions was the same as that for a single, unconjugated mSiNC (Fig. 6 E), and in all cases, the distributions could be fitted well with a single log-normal function (see the legend to Fig. 6 E) very similar to that shown in Fig. 4 B, showing that one mSiNC particle contributes to each mSiNC-Tf fluorescent spot.

These observations, together with the gel filtration HPLC results (Fig. 6, C and D), indicate that Tf precisely conjugated with
Figure 6. Conjugation of mSiNCs to Tf. (A and B) The scheme for conjugating mSiNC to Tf, using (N-c-maleimidocaproyloxy)sulfosuccinimide ester (sulfo-EMCS). (C, top) Superdex 200 gel filtration HPLC chromatograms for the samples obtained at 0 and 60 min after the mSiNC-Tf conjugation reaction was initiated. Peak 1, maleimide-Tf dimers (but not those cross-linked by mSiNC). Peak 2, overlapped eluent of mSiNC-Tf and maleimide-Tf. Peak 3, unconjugated mSiNC. The explanation for these assignments is given in the text. (bottom) Superdex 200 elution pattern of Tf monomers, dimers, and oligomers, generated by chemical cross-linking of Tf with bis(sulfosuccinimidyl)suberate. (D) Superdex 75 gel filtration HPLC chromatograms for the samples obtained at 0, 30, and 60 min after the mSiNC-Tf conjugation reaction was initiated. Peak 1, mSiNC-conjugated Tf (mSiNC-Tf). Peak 2, maleimide-Tf. Peak 3, unconjugated mSiNCs. The explanation for these assignments is given in the text. Additionally, these chromatograms indicate that about half of the mSiNCs added for this conjugation reaction became conjugated to maleimide-Tf in 60 min. Note that all of these peaks occur after the void volume determined by Blue Dextran 2000. Data in early time periods were deleted in C and D because no eluent peaks were detectable during these early periods. (E) Distributions of fluorescence intensities of individual mSiNC or mSiNC-Tf spots (measured in 530 × 530-nm areas) after the conjugation reactions for 0, 30, and 60 min. Excitation was by the 405-nm line of a solid-state laser. A single log-normal function (red curves) gave a good fit, and the two-component fitting did not significantly improve the fit, but nevertheless, the latter gave an estimate of the possible dimer fraction of less than a few percentages. Typical results among 7 (C, top), 3 (C, bottom), 11 (D), and 6 (E) entirely independent experiments are shown. a.u., arbitrary unit.
a single mSiNC was prepared and that mSiNC-Tf is monodisperse. Labeling of a protein with a nanocrystal at the 1:1 ratio with this precision now makes high-definition single-molecule tracking possible.

**Single-molecule tracking of Tf receptor (TfR) on the plasma membrane with mSiNC-Tf**

Each individual mSiNC-Tf bound to the bottom surface of the normal rat kidney (NRK) cell was observed with a TIRF microscope (both Cy3-labeled Tf [Cy3-Tf] and mSiNC-Tf readily enter the gap between the bottom membrane and the coverslip, practically reaching the equilibrium binding within a minute; Kasai et al., 2011; Fig. 7 A) at 37°C. The specific binding of mSiNC-Tf to TfR in the plasma membrane was confirmed by the following four experiments. (1) The mSiNCs that had not been conjugated to Tf did not bind to the cultured NRK cells (<1/100). (2) The inclusion of 100 nM free Tf in the incubation medium containing mSiNC-Tf strongly (~90%) inhibited mSiNC-Tf binding to the cell surface, suggesting that ~10% of mSiNC-Tf bound to the cell surface is nonspecifically adsorbed, consistent with the data of colloidal gold–conjugated molecules (~95%; Kusumi et al., 1993). (3) The mSiNC-Tf developed here, using human Tf, did not bind to the surface of chick embryonic fibroblasts (CEFs; Fig. 7 B). Because it has been shown that human Tf does not bind to chick TfR (Jing et al., 1990), this lack of binding further confirmed the specificity of mSiNC-Tf binding to TfR molecules on NRK cells (human Tf binds to rat TfR; Jing et al., 1990). (4) When mSiNC-Tf was applied to the culture of CEFs transfected with the GFP–human TfR (hTfR) gene, mSiNC-Tf bound only to the cells that expressed GFP-hTfR protein (the signal looks weak because the general staining of the cells was observed by epifluorescence microscopy; Fig. S5 A).

The single TfR–mSiNC-Tf complex could be tracked at video rate, without blinking or photobleaching, for ≤3,600 frames or 120 s (Fig. 7 C). Photobleaching prevented Cy3-Tf from being tracked for longer than 150 frames (5 s; Fig. 7 D), whereas blinking prevented the tracking of GFP-tagged molecules longer than 20 frames (~0.67 s; both in the case in which no reconnection protocol is used; see the next paragraph), indicating improvements by one to two orders of magnitude.

In the majority of single fluorescent molecule–tracking experiments reported in literature, even if the fluorescence signal from the probe disappears in several or many consecutive video frames (gap), the two trajectories separated by a gap are often “reconnected” into a single trajectory, assuming that the disappearance of the signal is caused by blinking (Table S2). This...
is how trajectories longer than the blinking time were obtained with the use of GFP-type molecules, QDs, and some organic dyes, such as Alexa Fluor 488 and Alexa Fluor 594.

In the present experiment, we aborted the tracking if we missed the observed spot even for a single video frame, i.e., no reconnection was performed. Even under such rigorous conditions, we were able to track single TIR molecules, tagged with mSiNC-Tf, for up to \( \approx 3,600 \) frames (120 s). This is about twice as long as the longest observation frames ever made using extensive reconnection protocols (Table S2). Note that, because the overall observation durations can be prolonged by time lapse imaging, the number of observed frames, and not the observed time period, is the critical parameter.

The failure to detect a fluorescent spot of an mSiNC particle in a single (or more) frames appears to occur for the following reasons: (a) temporary blurring of the fluorescent spots, as a result of z-directional motion, (b) release of the mSiNC-Tf from the receptor, (c) normal internalization processes of TIR molecules, and (d) long-term instability of the focus at 37°C, raised from the room temperature. With the use of mSiNCs, photobleaching and blinking are no longer the limiting factors for the durations of single-molecule tracking in living cells. Phototoxicity of the 440-nm laser illumination to the cell was not apparent, at least for 5 min (see Cell preparation for fluorescence microscopy in Materials and methods).

A statistical analysis (Kusumi et al., 1993) indicated that virtually all of the TIR molecules probed with mSiNC-Tfs at video rate underwent (apparent) simple Brownian diffusion (Fujiwara et al., 2002). The distribution of the effective diffusion coefficients in the time scale of 100 ms (Kusumi et al., 1993) is shown in Fig. 7 E. The median diffusion coefficient was 0.28 \( \mu m^2/s \) (n = 102), which is comparable with that found with single Tf molecules tagged with Cy3 (median, 0.26 \( \mu m^2/s \); n = 110; insignificant statistical difference). These results suggest that mSiNC-Tfs are bound 1:1 to the TIR molecules without cross-linking them, and mSiNC-Tfs themselves are not clustered (otherwise, mSiNC-Tf would have shown much slower diffusion by oligomerization-induced trapping; Iino et al., 2001), thus allowing the accurate observation of single-molecule diffusion of TIR.

Another representative type of the trajectories is shown in Fig. S5 B. After undergoing simple Brownian diffusion for 12.4 s, this TIR molecule underwent temporary immobilization for 1.7 s, as detected by a computer program (Fig. S5 C; Simson et al., 1995), and then disappeared from the TIR image (Fig. S5 B), suggesting the entrapment of TIR–mSiNC-Tf complex in a clathrin-coated pit, followed by internalization (Video 3). Previously, direct observation of a single molecule’s diffusion in the general plasma membrane area followed by its entrapment and internalization had never been achieved as a result of photobleaching in the case of organic dyes and GFP and perhaps because of slowed internalization of the receptor linked to a large particle, such as QDs and colloidal gold particles. Such direct observations of single receptor molecules undergoing diffusion in the plasma membrane and the subsequent internalization will further advance the studies about the mechanism of liganded receptor internalization and signaling in the intracellular membrane systems.

### Spatial variations of the TIR diffusion coefficient on the cell surface

Spatial variations of the physical properties of the plasma membrane have long been suspected. In the context of signal transduction function of the plasma membrane, spatial variations might be useful for creating space-dependent functional specializations, leading to, for example, polarized responses of the cells and/or enhanced interactions of molecules in specific locations in the plasma membrane (Chung et al., 2010). One of the useful parameters for describing the spatial variations is the diffusion coefficient of molecules, which might reflect the spatial variations in the interaction between the actin-based membrane skeleton meshwork and the plasma membrane, the densities of raft domains in the plasma membrane, and the densities of the binding sites for membrane molecules under investigation on the cortical actin filaments. However, spatial variations of the diffusion coefficients have not been systematically investigated. Such spatial variations can now be investigated with high reliability using a long trajectory of each individual molecule, which continuously explores various domains in the plasma membrane, reporting spatial variations of its local diffusion coefficient.

To obtain the spatial variations of the local diffusion coefficient, we used the display shown in Fig. 8 B (top; for the trajectory shown in Fig. 8 A), in which cumulative square displacement (SD; CSD; the sum of the SD taken every 33 ms) is plotted against time, with a relationship of CSD = 4Dr. Namely, the local slope/4 gives the local diffusion coefficient. Therefore, the changes in diffusivity could be identified in the plot, as the time intervals with different slopes (Chung et al., 2010). In this analysis, we imposed 5 s as the minimum duration for a given diffusivity state, within which any possible change in diffusion dynamics is ignored because a small number of plots leads to increasing uncertainty in the observations (see Detection of transient confinement zones and time [area]-dependent changes of diffusion coefficient in Materials and methods). In short, the long-term (for this study >5 s) local diffusion coefficient for the specific time period could be directly obtained from this plot (Fig. 8 B, bottom). From these displays, we understand that, even during the overall simple Brownian diffusion period, single TIR molecules can exhibit periods (>5 s) when they diffused at much smaller rates (Fig. 8 B, blue and red regions) or greater rates (Fig. 8 B, green region) than the average diffusion rate in a domain of 1–2 \( \mu m \) in diameter. Our attempt to find any correlation of slow- or fast-diffusion domains with the features that appeared in the enhanced bright-field images was not successful.

To further quantitate this observation, the distribution of long-term (>5 s, but still local) diffusion coefficients of TIR was obtained (Fig. 8 C, open bars). This distribution is compared with that for the diffusion coefficient determined on a 100-ms scale (Fig. 8 C, solid black bars). The average diffusion coefficient in the long-time regimen (median = 0.19 \( \mu m^2/s; n = 45 \)) is \( \approx 30\% \) smaller than that in the short-time regimen (\( \approx 100 \) ms; median = 0.28 \( \mu m^2/s; n = 102 \)), whereas the spread in the long-time regimen (a factor of several hundred, SD = 0.27 \( \mu m^2/s \)) is much greater than that in the short-time regimen (a factor of ~20; SD = 0.16 \( \mu m^2/s \)), as shown in Fig. 8 C. These
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results can be explained in the following way. TfR molecules crisscross at faster rates (and thus the diffusion coefficient in the 100-ms regimen is greater) without undergoing long-range diffusion (staying in a region of ~200 nm or less), probably confined within one or two compartments made by the partitioning by the actin-based membrane skeleton (fences) and its associated transmembrane proteins (pickets) or linked to the actin-based membrane skeleton that is undergoing rapid thermal fluctuation (Fujiwara et al., 2002; Kusumi et al., 2005), which could not be resolved at video rate (33 ms). Meanwhile, in the long-time regimen (>5 s) in which the macroscopic long-range diffusion matters, the diffusion of TfR appears slower because it experiences various diffusion barriers and temporary bindings during this period (but again each event could not be resolved at this time resolution and thus appears as decreases in the mean long-term diffusion coefficient), and the long-term diffusion coefficients vary greatly because, during the periods of >5 s, TfR molecules would experience many barriers, obstacles, and traps that vary greatly in numbers and properties depending on the location (micrometer level) of the TfR molecule in the plasma membrane. To the best of our knowledge, this is the first time that the diffusion coefficient in the time scale of 5 s or longer was measured for each individual molecule and was shown to spatially vary greatly (for exactly the same molecule), as much as a factor of several hundred.

Discussion

The problems of photobleaching, blinking, and cross-linking that fluorescent probes often exhibit are particularly severe in the studies using living cells, in which reducing–oxidizing chemicals cannot be used because of their toxic side effects, molecular oxygen cannot be removed, and the effects of cross-linking are severe. The mSiNCs developed in this study were subjected to most stringent tests of single-molecule imaging as well as many other characterizations and found to simultaneously solve many problems generally associated with fluorescent particles, dyes, and proteins. mSiNCs neither bleach nor blink at least for the total observation period of 300 min (540,000 frames) and are small (uniformly ~4 nm including the coat; although this size is greater than those of organic dye molecules) and monodisperse; they can be treated as water-soluble, monodispersed particles like highly soluble proteins, and a 1:1 conjugation of an mSiNC particle to a protein molecule was realized. When labeled with single mSiNCs, single receptors in the plasma membrane of living cells could be tracked for ~3,600 image frames, without missing it even for a single frame. This limitation was not caused by either blinking or photobleaching of the mSiNC. Phototoxicity of the 440-nm laser illumination to living cells was not detectable at least for 5 min in single mSiNC particle

Figure 8. Large variations of the long-term (>5 s) diffusion coefficient experienced by single molecules of TfR on the cell surface. [A] A typical trajectory of a TfR molecule tagged by mSiNC-Tf, recorded at video rate (33 ms/frame). Different colors indicate the portions of the trajectory with different diffusivity, as determined by the method described in B. [B] Quantitative analysis results of the trajectory shown in A. [top] The CSD plotted as a function of time. The plot is divided into subintervals, in which a single slope can describe the plot, by eye, and if the portion is longer than 5 s, its slope was obtained, as plotted at the bottom. [C] Distribution of the long-term (>5 s) diffusion coefficients, indicating the presence of long-range spatial variations in the membrane structure, probably reflecting local

variations in the actin-based membrane skeleton [open bars; obtained for 30 TfR molecules; 45 separate subintervals exhibiting different diffusion coefficients in time]. This should be compared with the histogram of 100-ms-scale diffusion coefficients (black bars, n = 102). The numbers in the box indicate median values (see arrowheads).
tracking. Because mSiNCs pass the rigorous tests for single-molecule tracking, they will be useful for general fluorescence imaging of live and fixed cells.

Both QDs and mSiNCs have their advantages and disadvantages. We envisage that mSiNCs will revolutionize single-molecule tracking in living cells. Meanwhile, for a few minutes tracking of membrane-bound receptors that diffuse slowly or are immobilized, blinking might not strongly affect the tracking, and therefore, QDs might be preferable as a result of more color selections (particularly for multicolor tracking) and short-term brightness. The availability of two different types of fluorescent biocompatible nanoparticles would be good for the fluorescence imaging community.

Using mSiNC-Tf, we were able to specifically label TIR and tracked the movement of the TIR–mSiNC-Tf complex on the NRK cell surface. Using mSiNC-Tf (1:1 conjugation) allowed one for the first time ever to directly observe, at the level of single receptor molecules, the initial entrapment of TIR within a clathrin-coated pit and the subsequent internalization process of each individual molecule. This was made possible by mSiNC because it does not photobleach or blink and is sufficiently small to allow internalization at rates comparable to physiological ligands. Therefore, mSiNC-conjugated ligands will make useful fluorescent markers to investigate their receptors’ internalization mechanisms by way of clathrin-coated pits and possibly of other domains, such as caveolae and GEECs (Chadda et al., 2007).

Furthermore, spatial variations of the diffusion coefficients of molecules in the plasma membrane, which have been sought after for a long time, have been discovered by using TIR labeled with mSiNC-Tf. This result is very reliable because spatial variations were found by single molecules that explored large areas on the cell surface, which the development of mSiNC-Tf made possible. Single TIR molecules (TIR–mSiNC-Tf complexes) exhibited various diffusion coefficients in membrane domains of 1–2-µm diameter, exhibiting a higher level of domain mosaicity in the plasma membrane, in addition to raft domains (3–100 nm) and membrane compartmentalization (30–300 nm) provided by the membrane skeleton (fence) and its associated transmembrane proteins (pickets; Fujiwara et al., 2002; Kusumi et al., 2005). The micrometer-level spatial variations might be caused by those of the actin-based membrane skeleton meshwork (spatial variations in mesh sizes and actin filament dynamics) as well as to spatial variations of the densities of raft domains and transient TIR binding sites. Spatial variations of the diffusion coefficient in the scale of 1 µm or longer might be useful for inducing polarized signal transduction and local cytoskeletal reorganization upon stimulation. These will be clarified in the near future by tracking long-term movements of various receptors tagged by mSiNCs.

One obvious application of mSiNCs will be their use as fiducial markers in single-molecule localization microscopy. mSiNC can be readily excited at the 405-nm laser line used for photoactivation photoconversion, and its fluorescence is detectable in broad regions between 500 and 700 nm, where fluorescence signal detection for localization microscopy is generally conducted. Overall, in this research, we took advantage of the fluorescent SiNCs developed in physical and chemical laboratories and developed truly useful SiNC-fluorescent probes for cell-biological and biomedical research, particularly for single-molecule imaging and tracking.

Materials and methods

Preparation of mSiNC and mSiNC-labeled Tf

Preparation of SiNC on the wafer. All the procedures for preparing mSiNC were conducted in the fume hood, and the relative humidity of the room was kept at <65%. All reagents, including HF, HNO3, acetic acid, and methanol, were of the reagent grade or better and were used without further purification and/or dehydration. A single-crystal silicon wafer (1100) orientation, 1–10 0-cm resistivity, and p-type boron doped; Mitsubishi Materials) was broken into smaller pieces (approximately 3.5 x 4.0 cm; Fig. 1 A, left). Etching was performed by immersing one side of the piece ~2 mm deep in HF/HNO3 (1:1, vol/vol) along the surface of the wafer (Fig. 1 A, second from the left) for a few seconds, to generate starting points for etching (preetching). The entire piece was then immersed in a HF/HNO3/H2O solution (2:7:8, vol/vol/vol) for 5 min (Fig. 1 A, third image). This etched wafer (wSiNC) was washed thoroughly in water purified by an ion-exchange resin and a reverse osmosis system (Milli-Q; EMD Millipore) and then was dried under vacuum (~0.1 mm Hg).

Surface silanization of the SiNC on the wafer (wSiNC) with mercaptosilane. Silanization was performed immediately after the etched wafers were dried (Fig. 1 B). Each wafer was submerged for 1.5 h in an acidic methanol solution (0.5 ml of 12 M HCl acid mixed with 500 ml methanol) containing 1% (vol/vol) 3-mercaptopropyltrimethoxysilane (Chisso). In this solution, 3-mercaptopropyltrimethoxysilane is hydrolyzed to produce 3MPTS. After silanization, the wafers were washed thoroughly in methanol and then were dried under vacuum (~0.1 mm Hg).

Recovery and purification of mSiNC. Nanocrystals were liberated from the polished side of the wafers by mechanical scraping of the silanized-etched surface using a razor blade and were suspended in water (mSiNC; Fig. 1 C). The suspension was sonicated for 2 h and was centrifuged at 12,000 g for 30 min. The mSiNC in the supernatant was further purified by an HPLC system (Hitachi) with a Superdex 75 column 10/300 GL (GE Healthcare) equilibrated in 50 mM phosphate buffer, pH 7.0, at a flow rate of 0.4 ml/min. Elution profiles were monitored by recording the absorption at 280 nm.

In a single preparation of mSiNC, we typically obtained ~2 nmol (typically ~1 µM x 2 ml) of mSiNC from a 6-inch wafer. This is comparable to a single commercial vial of Qdot655, which contains ~2 nmol (typically ~8 µM x 0.25 ml) of Qdot655. A project for scaling up the quantity of mSiNC in each preparation is in progress in our laboratories, and at the level of a basic science laboratory, a 30-fold increase appears to be achievable. Therefore, in the near future, we should be able to provide interested researchers with mSiNC for testing its applicability to their own projects. However, for the preparations at the industrial level, greater efforts would be required.

Preparation of mSiNC-labeled Tf (mSiNC-Tf). Tf was labeled with mSiNC, following the procedure illustrated in Fig. 6 (A and B). For the production of maleimide-Tf (Fig. 6 A, 20 µl N-[4-maleimidocaproyloxy)sulfo-succinimidyl ester (sulfo-EMCS; 10 mM; Thermo Fisher Scientific) in PBS, pH 7.2, was added to 200 µl Tf (holo-Tf, human; 0.1 mM; Sigma-Aldrich) in PBS, pH 7.2, and the mixture was incubated at room temperature for 60 min. Sulfo-EMCS–conjugated Tf (maleimide-Tf) was separated by chromatography on a Sephadex G-25 (GE Healthcare) column equilibrated with PBS. The maleimide/Tf molar ratio was adjusted to be low, i.e., 0.41 ± 0.05, using maleimide (sulfo-EMCS; 0.1 M). Maleimide/Tf concentration in the maleimide-Tf solution was determined in the following way. 40 µl mercaptoethylamine [0.125 M in 50 mM EDTA, pH 6.0] was first mixed with 100 µl of purified maleimide-Tf [0.01 mM], and the mixture was incubated at 30°C for 30 min, and then, the reduction of the thiol group in the mixture was determined by a reaction with 10 µl 4,4′-dithiodiipyridine [5 mM; at 30°C for 15 min; Grassetti and Murray, 1967] using the mercaptoethylamine reacted [room temperature for 60 min] with 1–64 µM sulfo-EMCS preincubated in water for 3 h to hydrolyze the succinimide group as standard solutions. The reaction was monitored by the optical density at 324 nm (ε = 19,800 M−1 cm−1). Assuming the Poisson distribution with a mean value of 0.41 maleimide group/Tf, ~6% of Tf should be conjugated with two or more maleimide groups.

To 40 µl of 10-µM maleimide-Tf in PBS, pH 7.2, we added 200 µl of 1-µM mSiNC and incubated the mixture at room temperature for 30 and
60 min. Tf labeled with mSiNC was separated by gel filtration HPLC [Superdex 75 and Superdex 200 columns (GE Healthcare)] equilibrated with 50 mM phosphate buffer, pH 7.0) from the nonlabeled Tf and the free mSiNC [Fig. 6, C–E]. In this reaction, the maleimide-Tf/mSiNC ratio in the reaction mixture was ~0.82 (because maleimide/Tf ratio is 0.41 and Tf/mSiNC ratio is 2). As seen in the peaks 2 and 3 in Fig. 6 D, ~20% of maleimide-Tf + nonlabeled Tf, i.e., ~50% of maleimide-Tf and ~40% of mSiNC, were consumed to form mSiNC-Tf within 60 min, providing a mean mSiNC-Tf ratio of ~0.4. Assuming the Poisson distribution with a mean value of 0.4 Tf molecules/mSiNC particles, ~6% of mSiNC should be conjugated with two or more molecules of Tf. That is, despite the presence of many sulfhydryl groups on the mSiNC, we can achieve almost 1:1 conjugation of Tf to mSiNC, particularly after the HPLC purification [Fig. 6 D, peak 1]. mSiNC conjugated with multiple Tf molecules were not detectable in the chromatogram shown in Fig. 6 D probably because their peaks were broad, overlapped, and small.

Characterization of mSiNC and mSiNC-Tf

**Determining the surface coating of wmSiNC and mSiNC.** A suspension of ~4-nm-diameter mSiNC, purified by Superdex 75 gel filtration HPLC, was dialedyzed against 50 mM NH4Cl using 3,500 molecular weight cut-off dialysis cassettes (Slide-A-Lyzer, Thermo Fisher Scientific). The surface coatings of the wmSiNC (mercaptosilane-coated SiNC on the wafer) and the mSiNC (dispersed nanocrystals in the dialysis solution were placed on the KBr plate and dried under ambient conditions) were analyzed by a spectrometer (1,024 acquisitions for a single spectrum; FT-2000 FTIR; Varian). Back-ground spectra were obtained using uncoated SiCN on wafers (wSiNC) or in solution.

**Determining spectroscopic properties of mSiNC.** Absorption spectra and fluorescence emission spectra were obtained with a spectrometer (U-3310; Hitachi) and a spectrophotometer (F-2500; Hitachi), respectively. The extinction coefficient of the mSiNC was determined from the absorption spectrum, and the concentration was determined by FCS measurements (described in subsection FCS and determination of the mSiNC size based on the FCS analysis). The correction for nonfluorescent crystals was made as described in Fig. 5, Table S1, and their related text (in the subsection Each in this reaction). The amount of mercaptosilane bound to, and the extinction coefficient of, mSiNC. A suspension of ~3.2-nm-diameter mSiNC, purified by gel filtration HPLC, was dialedyzed against 50 mM NH4Cl using 3,500 molecular weight cut-off dialysis cassettes (Slide-A-Lyzer). The resulting solution was freeze dried to remove water, NH4+, and CO2, and the remaining mSiNCs were weighed with a 100-ng scale balance [SE2; Sartorius].

**DLS measurement of the mSiNC size.** DLS of mSiNC purified by Superdex 75 gel filtration HPLC was measured using a particle size instrument (Zetasizer Nano ZS 90; Malvern Instruments Limited). The results were analyzed by its software, and the average hydrodynamic diameter (z average) of the mSiNC was determined using 3,500 molecular weight cut-off dialysis cassettes (Slide-A-Lyzer).

**FCS and determination of the mSiNC size based on the FCS measurements.** The FCS system (ConfoCor 3; Carl Zeiss) consisted of an inverted FluoTime 300 microscope equipped with a water immersion objective lens (40×, 1.2 NA; Carl Zeiss), an Ar+ laser, an avalanche photodiode, and a digital correlator. All samples were prepared with chambered 140-µm-thick cover glasses (Lab-Tek, Thermo Fisher Scientific) and were measured at 25°C. Samples were excited by the 458-nm line of the laser, and the fluorescence from the solution was separated by a dichroic mirror (458 nm) and a long-pass filter (470 nm), was detected.

Fluorescence autocorrelation curves G(τ) were fitted to a single-exponential diffusion model (Yoshida et al., 2001; Jin et al., 2007) as follows:

\[
G(\tau) = \left[ 1 + \left( \frac{\tau}{\tau_{\text{diffusion}}} \right) \left( \frac{\varepsilon_2}{\varepsilon_1} \right) \right]^{-1/2} \tag{1}
\]

in which N is the average number of fluorescent particles in the confocal volume of the lateral radius ωc and axial radius ωz, and τ_{diffusion} is the diffusion time of the fluorescent particles, which is related to the diffusion coefficient D and ωz by the following equation: τ_{diffusion} = ωz^2 / (4D).

For spherical particles, D is inversely proportional to its hydrodynamic radius r according to the Stokes–Einstein relationship (D = k_BT / 6πηr), in which k_B is the Boltzmann constant, T is the absolute temperature, and η is the viscosity of the solution (water = 0.894 cP at 25°C). Thus, the hydrodynamic diameters d_{H} of the mSiNC can be calculated by the following equation:

\[
d_{H} = \frac{6D}{\pi\eta} \left( \frac{\varepsilon_2}{\varepsilon_1} \right) \tag{2}
\]

where D is the diffusion coefficient of a standard dye, and τ_{diffusion} (mSiNC) and τ_{diffusion} (standard) are the diffusion times of the mSiNC and the standard dye, respectively. As the standard dye, rhodamine 6G (2.8 × 10^-6 cm^2/s; Rigler et al., 1993), monomeric GFP, and Qdot655 were used. Samples were measured for 10 runs of 20 s each at 25°C.

** TEM for determining the size distribution of mSiNC.** Purified mSiNC were mounted on polyvinyl Formvar-coated grids (Pelco) and imaged with a transmission electron microscope (JEM-1400 [JEOL] and Tecnai G2 Spirit [FEI]). The sizes of the individual mSiNC particles were determined using image software (Threshold: Maximum entropy and Analyze particles; National Institutes of Health).

**Determination of the amount of mercaptosilane bound to, and the extinction coefficient of, mSiNC.** A suspension of ~3.2-nm-diameter mSiNC, purified by gel filtration HPLC, was dialedyzed against 50 mM NH4Cl using 3,500 molecular weight cut-off dialysis cassettes (Slide-A-Lyzer). The resulting solution was freeze dried to remove water, NH4+, and CO2, and the remaining mSiNCs were weighed with a 100-ng scale balance (SE2; Sartorius).

The amount of mercaptosilane bound to mSiNC was determined by quantifying the thiols groups of 3-MPTS on the particle, by a reaction with 4,4-dithiodipyridine (Grassetti and Murray, 1967), using 1–4 µM 3-MPTS dissolved in purified water as standard solutions. A 100-µl sample was mixed with 40 µl of 50-mM EDTA, pH 6.0, and then incubated at 30°C for 30 min. We then added 10 µl of 5-MM 4,4-dithiodipyridine and incubated at 30°C for 15 min. The reaction was monitored by the optical density at 324 nm, and the mole amount of 3-MPTS (molecular weight = 155) bound to mSiNC was determined.

Next, by subtracting the weight of 3-MPTS bound to the mSiNC from the weight of mSiNC measured by a sensitive balance (as described in the first paragraph in this subsection), the weight of the mSiNC without the coating [i.e., SiNC] was determined. To obtain the molar amounts of mSiNC, the molecular weight of 3.2-nm SiNC (the diameter evaluated by EM) was evaluated in the following way. Because a 1-nm-diameter SiNC has a molecular weight of 836 [Si_24H_24], the molecular weight of the 3.2-nm SiNC can be 29 = 950.3 Si atoms and the 2D packing of hydrogen atoms on the surface ([3.2 nm/1 nm]_2 × 24 = 245.3 H atoms; the number of hydroxyl groups formed on the surface was not measured). Using this molecular weight, the mole amounts of 3.2-nm SiNC in given solutions (equal to the mole amounts of mSiNC in given solutions) were obtained.

These measurements allowed us to determine that 51.2 ± 19.4 (mean ± SD, n = 3) 3-MPTS molecules were bound to each mSiNC particle, giving a mean molecular weight of mSiNC of 36,083.6 (26,853.4 + 155 × 2). The extinction coefficient of mSiNC was determined from the absorption spectrum of a given solution and the mole amount of mSiNC in the solution.

**Long-term storage of mSiNCs and the preparation and observation of mSiNC-Tf in aqueous solutions during a day.** For long-term storage of the mSiNCs, they were stored in dehydrated benzene at 4°C for ≤60 d (Fig. S1). How ever, in aqueous solutions, the fluorescence intensity was reduced much faster. The fluorescence intensity of mSiNC decreases with an exponential decay time of 42 ± 2.8 h in 37°C, as described in Fig. S2. In a typical schedule of mSiNC conjugation to Tf and single-molecule observations of mSiNC bound to the cell surface (after obtaining the mSiNCs or after the change of the solvent from benzene to an aqueous buffer), it takes ~2 h for Tf conjugation and mSiNC-Tf purification and another 4 h for the single-molecule tracking observations for the day. Therefore, we examined the fluorescence spectrum, gel filtration HPLC elution pattern, and the FCS time series and autocorrelation function after incubation of mSiNC-Tf in
PBS at 37°C for 6 h. Based on these observations, we recommend that mSiNCs be kept in dehydrated benzene at 4°C for long-term storage and that on the day of the actual cell biology experiment, the benzene be evaporated by a flow of nitrogen gas flow and then aqueous buffer be added to dried mSiNCs.

Single fluorescent molecule imaging/tracking of mSiNC and mSiNC-labeled Tf

Single-color single fluorescent molecule tracking, using TIRF microscopy. Home-built objective lens-type TIRF microscopes were constructed on the basis of inverted microscopes (IX70 [Olympus] and TE2000 [Nikon]), equipped with a high immersion objective lens (Plan Apochromat 100×, NA 1.45 [Olympus]; Plan Apochromat 100× NA 1.49 [Nikon]) and 495-nm dichroic mirrors (FF562-DI02; Semrock), as described previously (Murakoshi et al., 2004; Koyama-Honda et al., 2005; Umemura et al., 2008). The 405- and 440-nm lines of solid-state lasers (BCL-050-405 and BCL-040-440; Crystalaser) were used to excite the mSiNC, Qdot655, and Cy3 samples for the Olympus- and Nikon-based stations, respectively. Stray excitation light was blocked with a long-pass filter (ES590LPv2; Chroma Technology Corp.). Fluorescence images were detected by an image intensifier (C8600-03; Hamamatsu Photonics) lens-coupled to an electron bombardment charge-coupled device (CCD) camera (C7190-23; Hamamatsu Photonics), and the obtained images were recorded on a digital videotape (PDV-184AME; Sony). Each individual fluorescent spot was identified and analyzed by using a homemade computer program as described previously (Fujiwara et al., 2002). All of the observations were made at 37°C using Hanks’ buffered salt solution.

The TIRF apparatus is not required for the observations of single mSiNC particles, and a typical epifluorescence apparatus is sufficient for single mSiNC particle tracking at video rate. However, because our focus in this research is single fluorescent molecule (particle) tracking and because TIRF illumination is now a standard in laboratories conducting single fluorescent molecule tracking, we also show the data obtained by TIRF illumination in this study.

Two-color (two molecular species) single fluorescent molecule tracking by TIRF microscopy. An objective lens-type TIRF microscope was constructed on an inverted microscope (TE2000; Nikon), equipped with an oil immersion objective lens (Plan Apochromat 100× NA 1.49; Nikon) and 495-nm dichroic mirrors (FF562-DI02; Semrock), as described previously (Murakoshi et al., 2004; Koyama-Honda et al., 2005). The 440 (BCL-040-440; Crystalaser) and 488 (Excelsior One-488-100; Spectro-Physics)-nm lines of solid-state lasers were used to excite the Alexa Fluor 488-mSiNC samples.

The fluorescence images of mSiNC and Alexa Fluor 488 were separated by a dichroic mirror (FF562-DI02; Semrock) and projected into two detection arms with a long-pass filter (>590 nm for mSiNC; ES590LPv2) and a band-pass filter (520/35 nm for Alexa Fluor 488; Semrock). The fluorescence images were recorded and analyzed as described in the previous section.

Fluorescence intensity measurements. Fluorescence images were digitized frame by frame with a homemade computer program (Fujiwara et al., 2002). The signal intensity of 330 × 330-nm areas (8-bit images in 10 × 10 pixels), each containing a single spot, was measured. The background intensity of an adjacent area was subtracted for each spot.

Detection of transient confinement zones and time (area)-dependent changes of diffusion coefficient. To detect nonrandom confinement in a zone, or a transient confinement zone, we looked for periods in which the protein remains in a membrane region for a duration considerably longer than a Brownian diffusant would stay in an equally sized region, based on the results of 500 simulated Brownian trajectories, the probability level remains in a membrane region for a duration considerably longer than a random diffusant would stay in an equally sized region, based on the theory and program developed by Simson et al. (1995). For every point within a trajectory, the probability level was equilibrated with 50 mM phosphate buffer, pH 7.0. The dye/mSiNC ratio was determined by linearly fitting the plot of MSD versus the time in-crement. D100ms, corresponding to Dc, defined in Kusumi et al. (1993), was determined by using their re-pective molar extinction coefficient of 53,000 and 71,000 M⁻¹ cm⁻¹ [at 400 and 495 nm, respectively (Table 5)]. There is virtually no overlap of the spectra of mSiNC and Alexa Fluor 488 at these wavelengths.

Using Alexa Fluor 488–labeled mSiNCs [Alexa Fluor 488–mSiNC] and determination of the fraction of mSiNCs that are actually fluorescent. The Alexa Fluor 488–labeled mSiNCs was basically prepared essentially as described previously (Fujiwara et al., 2002; Umemura et al., 2008). In brief, 10 µl Alexa Fluor 488 C5-maleimide dye (0.2 mM; Molecular Probes) in dimethylformamide was added to 200 µl mSiNC (~1 µM) in 20 mM phosphate buffer, pH 7.0, and the mixture was incubated at room temperature for 60 min. Alexa Fluor 488–mSiNC (unreacted mSiNC) was separated from the free Alexa Fluor 488 dye by Superdex 75 gel filtration HPLC equilibrated with 50 mM phosphate buffer, pH 7.0. The dye/mSiNC ratio of the samples used for two-color single fluorescent molecule tracking was 0.35 ± 0.031 (mean ± SD; n = 3), which was determined using their respective molar extinction coefficient of 53,000 and 71,000 M⁻¹ cm⁻¹ [at 400 and 495 nm, respectively (Table 5)]. There is virtually no overlap of the spectra of mSiNC and Alexa Fluor 488 at these wavelengths.

Using Alexa Fluor 488–labeled mSiNCs at a mean Alexa Fluor 488/MsiNC molar ratio of 0.35, we examined the fractions of fluorescent and nonfluorescent mSiNCs. For this examination, the fluorescence signals emitted from Alexa Fluor 488 and mSiNCs, each excited with a 488- or 440-nm laser, were observed (Fig. 6). Alexa Fluor 488 signals in the view field were identified because of possible FRET from Alexa Fluor 488 to mSiNCs (although this FRET efficiency appeared to be very low because single-particle spots were hardly detected in mSiNC channel with the excitation at 488 nm), some of the Alexa Fluor molecules might not have been detected in the Alexa Fluor 488 channel, but this will not affect the final result because we made further examination only for the detected Alexa Fluor spots. Then, each identified Alexa Fluor spot was examined whether it was colocalized by an mSiNC spot (Koyama-Honda et al., 2005; No single-particle spots were detected in the Alexa Fluor 488 channel with the 440-nm excitation for mSiNC, indicating very low efficiency of FRET from mSiNC to Alexa Fluor488). This way, the fraction of Alexa Fluor 488 spots colocalized by fluorescent mSiNC spots was determined. The numbers of Alexa Fluor spots that were colocalized or noncolocalized by the mSiNC spots were counted (the Alexa Fluor 488/mSiNC ratio of 0.35 did not affect further
calculations of the percentage of dark mSiNC particles because we first detected Alexa Fluor 488 spots and then examined for each Alexa Fluor 488 spot whether it is colocalized by the mSiNC fluorescence spot; Fig. 5). The result showed that 38.4 ± 2.6% (mean ± SD) of the SiNCs were actually fluorescent (for six independently prepared specimens; examining 245 Alexa Fluor 488 spots in total), indicating much improvement from the previously reported value of 2.8%, although the improvement mechanism is not clear.

Preparation of Cy3-labeled Tf (Cy3-Tf). The Cy3-labeled Tf was basically prepared as described previously (Fujimaru et al., 2002; Umemura et al., 2008). To outline the process briefly, to 200 µl Tf (10 µM) in PBS, pH 7.2, 10 µl Cy3 monoactive dye (1.6 mM; GE Healthcare) in dimethylformamide was added, and the mixture was incubated at room temperature for 60 min. Cy3-Tf (=unlabeled Tf) was separated from the free Cy3 dye by chromatography on a Sephadex G-25 column (GE Healthcare) equilibrated with PBS. The dye/protein ratio of the samples used for single fluorescent molecule tracking was 0.39 ± 0.062 (mean ± SD; n = 4), using the molar extinction coefficients of 150,000 M⁻¹ cm⁻¹ (at 552 nm) for Cy3 (Table S1) and 83,360 M⁻¹ cm⁻¹ (at 280 nm) for Tf (West et al., 2000).

Cell preparation for fluorescence microscopy. To outline the process briefly, the NRK fibroblast cell line was grown in Ham’s F12 medium supplemented with antibiotics and 10% FBS. CEF cells, purchased from ATCC, were grown in DMEM medium supplemented with antibiotics, 5% tryptose phosphate broth, and 5% FBS. Cells were plated on 12-mm φ coverslip-based dishes (IWAKI) 2 d before being observed. Shortly after observation, the cells were preincubated in Hanks’ buffered salt solution at 37°C for 15 min to deplete Tf from the cell surface, and then, mSiNC-Tf or Cy3-Tf, eluted from the column, was added to a final concentration of 0.5–5 nM.

As shown in Fig. 7, the cells internalize TfRs (labeled with mSiNC-Tf) at the same rate, even after illumination with a 440-nm laser line, for ≥5 min. The cellular morphology did not change appreciably during this period. Therefore, although due caution is certainly required, our observation conditions of exciting mSiNC on the cell surface with a 440-nm laser line would be acceptable for the types of experiments described in this study.

Determining the observation durations for single particles/molecules. The observation duration for a single molecule (particle) was determined in terms of the number of observation frames [images], rather than in terms of the total observation time. Because the overall observation duration expressed in terms of time can be prolonged by time-lapping, the number of observed frames, and not the observed time period, is the critical parameter. Generally, in single-molecule (particle) tracking, even if some frames miss the signal from the tracked single molecule, researchers tend to connect the gap-separated trajectories into a single longer trajectory, and sometimes, the gap lengths are very long or undescribed in the study (Table S1) and 83,360 M⁻¹ cm⁻¹ (at 280 nm) for Tf (West et al., 2000).

Online supplemental material Fig. S1 shows that the fluorescence properties of the mSiNCs are not affected during storage for 60 d in dehydrated benzene. Fig. S2 shows time-dependent changes of the integrated fluorescence intensity (excited at 350 nm) of the mSiNC-Tf in 50 mM phosphate buffer, pH 7.0. Fig. S3 shows that mSiNC neither blinked nor photobleached, whereas QDs frequently blinked and gradually photobleached. Fig. S4 shows that fluorescence intensity of mSiNC is linearly increased with an increase of the excitation laser intensity at least up to 0.68 µW/µm², twice the intensity generally used in the present research. Fig. S5 shows specific binding of mSiNC-Tf [human Tf] to TfR and a typical trajectory of this molecular complex (mSiNC-Tf and TfR) on the NRK cell surface including a period of temporary confinement followed by a (probable) internalization process. Table S1 summarizes spectral properties of mSiNC, commercial organic dye, and Qdot655. Table S2 shows the total number of consecutive trajectories (frames) acquired for single particles of Qdot655, Cy3, and mSiNC before the particle became untrackable and their maximal gap in reconnection. Video 1 shows fluorescence images of single mSiNCs and single QDs. Video 2 shows that individual TfR molecules, each tagged with an mSiNC-Tf, undergo rapid apparent simple Brownian diffusion in the plasma membrane of a live NRK cell. Video 3 shows the result of direct visualization of internalization process of a single molecule tagged with mSiNC-Tf in the plasma membrane of a live NRK cell. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.201301053/DC1.

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Author contributions: H. Nishimura performed a large majority of the experiments reported here. K.P. Ritchie and M. Goto developed a new SiNC preparation method at the initial stages of this work. N. Morone performed TEM, and H. Nishimura and H. Sugimura performed FTR observations. Y. Nakano and J. Sase designed and produced the autofocus system for the microscope. H. Nishimura, R.S. Kasai, and T.K. Fujimaru performed singlemolecule imaging. A. Yoshimura, K. Tanaka, I. Sase, Y. Nakano, and A. Kusumi evaluated...


