Orally bioavailable glutamine antagonist prodrug JHU-083 penetrates mouse brain and suppresses the growth of MYC-driven medulloblastoma\(^1,2\)

**Abstract**

A subset of poor-prognosis medulloblastoma has genomic amplification of \(\text{MYC}\). \(\text{MYC}\) regulates glutamine metabolism in multiple cellular contexts. We modified the glutamine analog 6-diazo-5-oxo-l-norleucine (DON) to mask its carboxylate and amine functionalities, creating a prodrug termed JHU-083 with increased oral bioavailability. We hypothesized that this prodrug would kill MYC-expressing medulloblastoma. JHU-083 treatment caused decreased growth and increased apoptosis in human MYC-expressing medulloblastoma cell lines. We generated a mouse MYC-driven medulloblastoma model by transforming C57BL/6 mouse cerebellar stem and progenitor cells. When implanted into the brains of C57BL/6 mice, these cells formed large cell/anaplastic tumors that resembled aggressive medulloblastoma. A cell line derived from this model was sensitive to JHU-083 in vitro. Oral administration of JHU-038 led to the accumulation of micromolar concentrations of DON in the mouse brain. JHU-083 treatment significantly increased the survival of immune-competent animals bearing orthotopic tumors formed by the mouse cerebellar stem cell model as well as immune-deficient animals bearing orthotopic tumors formed by a human MYC-amplified medulloblastoma cell line. These data provide pre-clinical justification for the ongoing development and testing of orally bioavailable DON prodrugs for use in medulloblastoma patients.
**Introduction**

Medulloblastoma is the most common malignant brain tumor of childhood. Though 60% of medulloblastoma patients survive for at least 10 years, survival varies widely depending on the molecular genetics of the patient’s tumor [43]. RNA expression profiling and DNA methylation studies have subdivided medulloblastoma into at least four molecular subgroups: WNT, SHH, Group 3, and Group 4 [25,33]. The standard treatment for medulloblastoma is surgical resection of the tumor, radiation and chemotherapy. This therapy is associated with high morbidity; medulloblastoma patients often experience cognitive delays, learning disabilities, hearing loss, increased risk for future malignancies and stroke, endocrine dysfunction, and cerebellar mutism syndrome [5,15,24,39,44]. A subset of patients within Group 3 have amplification or over-expression of the MYC proto-oncogene. These patients have among the worst clinical outcomes of all medulloblastoma patients [2,7]. The relatively poor prognosis for many medulloblastoma patients and the severe complications faced by survivors indicate an urgent need for more effective and less toxic therapies.

MYC regulates a variety of cellular processes, including metabolism of both glucose and the amino acid glutamine [9]. Glutamine is the primary nitrogen source for synthesis of nucleic acids, other amino acids, and hexosamines. Glutamine can also be used as a carbon source to replenish TCA cycle intermediates, in a process called glutaminolysis [27,31]. Many types of MYC-driven cancers uptake large quantities of glutamine, some to the point of becoming “addicted” and being unable to survive without glutamine [10,12,26,47]. MYC regulates the utilization of glutamine by upregulating genes involved in glutamine uptake and metabolism, ASCT2, LAT-1 and glutaminase (GLS) [4,18].

The role of glutamine metabolism in MYC-driven medulloblastoma is underexplored, but a study by Wilson et al [46] supports the hypothesis that a subset of medulloblastoma tumors have elevated glutamine metabolism. Wilson et al non-invasively measured glutamate levels in the tumors of medulloblastoma patients using magnetic resonance spectroscopy (MRS). They found that patients with high-levels of glutamate in their tumors had much poorer survival than patients with low glutamate levels. The initial step in glutamine metabolism is the conversion of glutamine to glutamate by GLS. High levels of glutamate indicate that glutamine is being metabolized at a high level. Thus, increased glutamine metabolism is associated with poor-prognosis medulloblastoma. Because glutamine metabolism is a potential therapeutic target in other MYC-driven cancers [11,48], we hypothesized that targeting glutamine metabolism in MYC-driven medulloblastoma could be a potentially useful strategy in this devastating tumor type.

**Methods**

2.1 JHU-083

The synthesis of JHU-083 (Ethyl 2-(2-Amino-4-methylpentanamido)-DON) was conducted as detailed by our group previously [37]. For in vitro testing, JHU083 was dissolved in sterile water; for in vivo studies it was dissolved in sterile PBS buffer. JHU-083 doses in this paper are given as the DON-equivalent dose. Due to the presence of the promoiities, 1.83 mg of JHU-083 equals 1.0 mg of unmodified DON.

2.2 Cell Culture

The patient-derived medulloblastoma cell line D425MED was established at Duke University [21]. It is grown in MEM media supplemented with 10% FBS. We generated human neural stem cell models of MYC-driven medulloblastoma or SV40 immortalized control cells as described [20]. Our MYC-expressing models recapitulate Group 3, MYC amplified medulloblastoma by expression profiling and phenotype [20]. These human neural stem cell lines grow as neurospheres in "EF" media composed of 30% Ham’s F12, 70% DMEM, 1% antibiotic antymycotic, 2% B27 supplement, 5ug/mL heparin, 20 ng/mL EGF, and 20 ng/mL FGF2 [20]. Those harboring AKT are grown under puromycin selection. The MED211 patient derived xenograft was obtained from the Brain Tumor Resource Lab and is described in [6]. A cell line was derived from this PDX model by removing tumor tissue from tumor-bearing mice at the time of sacrifice and passing tumor through a P1000 pipette. Cells were grown in EF media. All cells were verified to be mycoplasma free by PCR testing. Cell line identity testing was performed by the Johns Hopkins Genetic Resources Core Facility and is included in Supplemental Figure 1.

2.3 Mouse Medulloblastoma Model

Creation of this model is described in detail in the main body of the manuscript. The cells are grown in EGF/FGF media made with a 50:50 mixture of F12:DMEM, but is otherwise identical to that described above. The R248W-TP53 plasmid (Addgene plasmid 16437) and MYC plasmid (Addgene plasmid 17758) were subcloned into pWPI (Addgene 12254) [29]. Lentivirus was produced by transfecting 293T cells with VSV-G envelope plasmid, Δ8.9 gag/pol plasmid and the plasmid containing the gene of interest as described in [36] using Fugene (Roche) per manufacturer’s instruction. The supernatant was collected at 48 and 72 h. The collected supernatant was filtered with a 0.45 micron filter and concentrated overnight at 4°C using 5% PEG 8000 and 150 mM NaCl then centrifuged for 30 minutes at 2000xg. Concentrated virus was stored at -80°C in serum free DMEM until use. Visualization of the distribution of this model in situ was provided by fluorescence imaging microscopy using a dual camera setup for GFP and white light (anatomical) imaging. The excised brain was illuminated by a SpectraX LED light source (Lumencor) with emission light captured by an F1.2 80 mm lens (Nikon), and filtered by a dichroic and low pass filter in an OptosplitII (Cairn) tube to dual PixelFly CCDs (PCO). Data was acquired in Metamorph NX.

2.4 Cell growth Assay

Experiments were performed in 96 well plates. Equal numbers of cells were plated in triplicate for each treatment. 20uL of CellTitre 96 Aqueous One Solution (Promega) was added to each well for every 100uL of media and incubated at 37°C for 1 hour. Absorbance was read at 490 nM on a plate reader. Growth between vehicle and drug treated cells was statistically analyzed using Student’s t-test.

2.5 Western blots

Protein was extracted from cell pellets using RIPA buffer and quantified using a Bradford Assay. Antibody against cleaved-PARP is from Cell Signaling technologies (#9541). Antibodies against ACTIN (sc-47778) and MYC (sc-40) are from Santa Cruz Biotechnologies. The following dilutions were used cleaved-PARP (1:800), ACTIN (1:1000), and MYC (1:1000). Peroxidase labeled secondary antibodies are from...
Cell Signaling Technologies and used at a 1:3500 dilution. Bands were quantified using ImageJ.

2.6 Cleaved Caspase-3 immunofluorescence
Following 72 h drug treatment, cells were fixed in cytospin fluid and spun onto glass slides using a cytocentrifuge. Cells were permeabilized in 0.1% TritonX, blocked in 5% normal goat serum, and incubated with anti-cleaved caspase-3 antibody diluted 1:400 (Cell Signaling Technology®, clone 5A1E), followed by a secondary antibody conjugated to Cy3 diluted 1:500 (Jackson Immunoresearch). DAPI was used as a counterstain. Three DAPI and the corresponding Cy3 images were taken for each slide. The number of DAPI and Cy3 positive cells were counted using Adobe® Photoshop®. For each pair of images, the percent of Cy3 positive cells was calculated. The average Cy3 positivity was determined by calculating the average of at least three pairs of images for each treatment. Images were blinded before counting. Vehicle and drug treated cells were compared by Student’s t-test.

2.7 Histology
Brains were fixed in 10% buffered formalin for a minimum of 24 h. The Johns Hopkins Pathology Reference Lab processed tissue for paraffin embedding and sectioning.

2.8 Immunohistochemistry
All IHC staining was performed by the Johns Hopkins Oncology Tissue services.

2.9 Animal Studies
Xenografting was conducted in female mice of a minimum of 6 weeks old. D425MED cells were injected into nude (Nu/Nu) mice. mNSC DNp53 MYC cells were injected into C57BL/6J mice. Prior to implantation of cells, animals were anesthetized using a mixture of 10% ketamine and 5% xyaline. A burr hole was made in the skull 1 mm to the right and 2 mm posterior of the lambdoid suture with an 18 gauge needle. The needle of a Hamilton syringe was inserted 2.5 mm into the brain using a needle guard and 100,000 D425MED cells or 50,000 mCB DNp53 MYC in 3uL of media were injected into the cerebellum. JHU-083 was prepared in sterile PBS and orally bioavailable and converts to DON after administration (Figure 1) [37] [49].

3.2 A cell-based mouse model of MYC-driven medulloblastoma
In addition to utilizing existing patient-derived medulloblastoma cell lines, we created a novel mouse model using mouse cerebellar stem and progenitor cells to facilitate in vivo studies in the context of an intact immune system (Figure 2A). The developing cerebellum was dissected out of E15 C57BL/6J fetuses, the tissue manually dissociated and placed in EGF/FGF containing, serum-free cell culture media. These cells grow as neurospheres. The isolated stem and progenitor cells were infected with lentivirus bearing human dominant-negative TP53 (DNp53) and human MYC, both of which are associated with aggressive, poor-prognosis medulloblastoma [7,33].

To generate subclones, individual neurospheres were manually selected with aid of a dissecting microscope and expanded before the expression of MYC and P53 was verified by western blot. A representative sub-clone (Figure 2B) shows robust expression of both MYC and TP53. Sub-clones positive for both oncogenes were injected into the cerebella of 4–6 week old adult female C57BL/6J mice. If a subclone formed a tumor, the tumor was implanted into additional mice. This in vivo passaging was repeated five times. A cell line was generated from the fifth passage. The tumors formed from mCB DNp53 MYC cells resemble human anaplastic medulloblastoma, with leptomingeal dissemination (Figure 2C, D) as well as a large cell/anaplastic phenotype (Figure 2E, F) [14]. These tumors also showed very robust expression of MYC (Figure 2G) and TP53 (2H) by IHC, similar to group 3 MYC-driven medulloblastoma [20].

3.3 JHU-083 reduces growth of MYC-expressing medulloblastoma cell lines
Treatment with 10uM of JHU-083 significantly reduced the growth of MYC-expressing medulloblastoma models as measured by MTS assay (Figure 3). All the cell models tested, except for human neural stem cells immortalized with SV40, express robust amounts of MYC (Figure 3A).

In the patient derived, MYC-amplified cell line D425MED, 10uM JHU-083 treatment decreased growth by 80% at day 5 of treatment

![Figure 1. Chemical structure of 6-diazo-5-oxo-l-norleucine (DON) and the prodrug JHU-083 (Ethyl 2-(2-Amino-4-methylpentanamido)-DON).](image-url)
In human cerebellar neural stem and progenitor cells transformed by MYC alone, 10μM JHU-083 treatment caused a 98% decrease in growth at day 7 of treatment (\(P = 5.2 \times 10^{-6}\)).

Human neural stem and progenitor cells transformed with dominant negative TP53, human telomerase (hTERT), AKT and MYC form tumors that phenocopy large cell/anaplastic human medulloblastoma when injected into the cerebella of nude mice. This model also mimics the expression profile of the C1 subgroup of medulloblastoma, the subgroup with the poorest prognosis that consists of MYC-expressing tumors [20]. In two subclones created by the addition of dominant-negative TP53, hTERT, constitutively-active AKT, and MYC to human cerebellar stem and progenitor cells, 10μM JHU-083 reduced growth at day 7 by 55% (\(P = 3.5 \times 10^{-5}\), Figure 3D) and by 50% in #5 (\(P = 0.03\), Figure 3E). JHU-083 treatment significantly decreased the growth of mouse cerebellar stem and progenitor cells transformed with MYC and dominant-negative TP53 (Figure 3F). 72 h treatment with 10μM JHU083 lead to an 86% reduction in growth by day 4 of treatment (\(P = 7.23 \times 10^{-6}\)).

3.4 JHU-083 treatment increases apoptosis in MYC-expressing medulloblastoma cell lines

Treatment with 10μM JHU-083 for 24 h caused an average 3.5-fold increase in the expression of cleaved-PARP as measured by Western blot in five MYC-expressing medulloblastoma cell lines (Figure 4A), indicating that JHU-083 treatment induces apoptotic cell death. Treatment with 10μM or 20μM of JHU-083 for 72 hours similarly caused a significant increase in the percentage of cleaved caspase-3 positive cells as measured by immunofluorescent staining (Figure 4B). In the patient-derived cell line D425MED, 10μM and 20μM of JHU-083 increased the percentage of cleaved caspase-3 positive cells from 3% to 14% (\(P = 1.99 \times 10^{-6}\)) and 29% (\(P = 1.9 \times 10^{-7}\)) respectively.

We generated the MED211 cell line from a medulloblastoma patient-derived xenograft (PDX) tumor. Genetic analysis indicates that this PDX has MYC amplification and belongs to the G3 medulloblastoma subgroup [6]. In MED211 cells, treatment with 10μM or 20μM JHU-083 increased the percentage of cleaved caspase-3 positive cells from a baseline level of 22% to 64% (\(P = 6.2 \times 10^{-11}\)) and 75% (\(P = 2.4 \times 10^{-11}\)). In human cerebellar neural stem and progenitor cells transformed with MYC alone, 10μM and 20μM of JHU-083 increased cleaved caspase-3 levels from 22% to 32% (\(P = 0.03\), Figure 3G) and then to 47% (\(P = 1.5 \times 10^{-5}\)). In CB DNp53 MYC #2, 10μM JHU-083 and 20μM JHU-083 increased cleaved caspase-3 levels from 8% to 22% (\(P = 8.2 \times 10^{-6}\)) and 41% (\(P = 9.7 \times 10^{-9}\)) respectively.

In the mouse medulloblastoma cell line mCB TP53 MYC, 10μM JHU-083 and 20μM JHU-083 increased cleaved caspase-3 levels from 12% to 30% (\(P = 0.0002\)) and 47% (\(P = 2.9 \times 10^{-5}\)). In the mouse medulloblastoma cell line mCB TP53 MYC, 10μM JHU-083 and 20μM JHU-083 increased cleaved caspase-3 levels from 8% to 22% (\(P = 8.2 \times 10^{-6}\)) and 41% (\(P = 9.7 \times 10^{-9}\)) respectively.
levels from 34% to 50% \((P=2.4\times10^{-6})\) and 59% \((P=1.5\times10^{-7})\). Non-MYC expressing human neural stem cells immortalized with SV40 were not sensitive to treatment with JHU-083 at 10uM or 20uM \((P>0.82)\). These results indicate that JHU-083 induces apoptosis in MYC-expressing medulloblastoma cell lines and has little effect on non-MYC expressing cells. To further address the mechanism of high-MYC cells being sensitive to JHU-083, we determined the expression of glutaminase \((GLS)\) in our neural stem cell models by western blot. GLS is regulated by MYC and is a key enzyme governing the conversion of glutamine to glutamate \([8]\). When cells transduced with SV40 do not express detectable MYC and do not express GLS, while cells transduced with MYC alone or additional oncogenic drivers such as R248WTP53 and activated AKT express robust levels of GLS (Supplemental Figure 2).

3.5 **JHU-083 administration achieves micromolar concentrations of DON in the brain**

To determine if DON delivered by JHU-083 is capable of crossing the blood–brain barrier, nude mice were administered JHU-083 by oral gavage (20 mg/kg). After 1 hour, the animals were euthanized and the brains removed. The cortex, cerebellum and brain stem were manually separated and flash frozen in liquid nitrogen. DON levels in the brain were subsequently quantified via LC–MS/MS as our laboratory has previously described \([49]\). There was no significant difference in the concentration of DON in the three regions of the brain analyzed \((P=0.42\), one-way ANOVA, \(n=5\) \(\text{Figure 5A}\)). The cerebellum had an average concentration of 11.3 \(\mu\text{M}\) of DON. Because 10uM JHU-083 successfully reduces growth of MYC-expressing medulloblastoma cell lines (\(\text{Figure 3}\)) and induces apoptosis (\(\text{Figure 4}\)), we hypothesized that a 20 mg/kg dose of JHU-083 would significantly extend the survival of animals bearing MYC-expressing medulloblastoma tumors.

3.6 **JHU-083 extends the survival of animals with MYC-driven medulloblastoma orthotopic xenografts**

Treatment with JHU-083 (20 mg/kg twice weekly) extended the survival of athymic nude mice bearing D425MED orthotopic xenografts from 21 to 28 days, an increase of 29 percent \((P = 0.006\) by Log-rank
Discussion

Patients with MYC-expressing medulloblastoma face a grim prognosis. There are currently no biologically informed therapies available for these patients. The experiments presented here show the potential for a metabolic approach to treating this tumor type. The majority of MYC-positive medulloblastoma tumors fall into the G3/C1 subgroup. Other studies have shown that many relapsed medulloblastoma tumors are positive for MYC expression upon relapse, regardless of subgroup [22], suggesting that targeting MYC-regulated metabolism could have utility in recurrent medulloblastoma of other subtypes as well as G3/C1 patients.

A critical aspect of this study is that it compares efficacy of a novel glutamine antagonist against MYC-driven medulloblastoma models of both human and mouse origin and demonstrates a therapeutic response in both immune deficient and immune competent CNS microenvironments. Our mCB DNp53 MYC cells are derived from C57BL/6J mice and are rapidly tumorigenic in immune intact animals. *TP53* mutations and *MYC* amplification are not only associated with poor outcomes in medulloblastoma at diagnosis, they are also the most frequently gained mutations in relapsed medulloblastoma and both are associated with aggressive disease phenotypes [22]. Since novel therapeutics will initially be tested in human patients with relapsed disease and intact immune systems, the mCB DNp53 MYC cells present an ideal system for modeling treatment effects in relapsed medulloblastoma.

This model differs from other similar models [35] in that it was established in an immune intact mouse, utilizes human oncogenes to mimic human tumor biology in a mouse cell of origin and in that lentiviral vectors were used to deliver these oncogenes. Since medulloblastoma is an immunologically “cold” tumor [30], lentivirus was used to decrease the risk of potentially immunogenic insertional mutagenesis that can occur with retroviral models [23]. Careful clone selection after in vivo adaptation has led to the generation of a stable cell line model of medulloblastoma available for orthotopic use in any transgenic mouse strain developed on the C57BL/6J background, one of the oldest and most widely used mouse strains in biomedical science.
We observe a similar survival effect of JHU-083 in our immuno-deficient and immune-intact medulloblastoma models. Although dissecting the effects of DON and DON prodrugs on the adaptive immune system is beyond the scope of this study, we include an immune-competent MYC-driven medulloblastoma model to demonstrate that a therapeutic benefit is preserved in this setting. The dependence of activated lymphocytes on glutamine is well described and reviewed recently by Bettencourt and Powell [3]. DON and JHU-083 have a marked immunosuppressive effect in models of cerebral malaria, decreasing edema and improving survival of afflicted mice, with corresponding inhibition of CD-8 T-cell degranulation [19,40]. DON similarly exerts robust anti-inflammatory effects a viral mouse model of encephalitis [28]. Interestingly, when DON treatment stopped, the inflammation resumed and rapidly progressed, leading to the death of mice [28]. The rebound inflammation seen in viral encephalitis suggests that clinical use of DON might, similar to high-dose cyclophosphamide, alter the microenvironment in ways that could be manipulated to enhance the immune response [17]. JHU-083 alters microglial metabolism by suppressing glutaminase activity [50]. Changes in microglial/macrophage metabolism may affect the polarity of these cells and alter the immune microenvironment, contributing to or thwarting an immune response [34].

These studies indicate that DON/JHU-083 has a profound impact on both innate and adaptive immunity. Demonstrating that the anti-tumor effects of JHU-083 are preserved in the presence of an intact immune system therefore lends important translational strength to our findings. Many preclinical studies are unable to include complementary data in immune intact systems due to a lack
of appropriate models, and this may represent an overall obstacle to clinical translation. Our immune-intact model presents an opportunity to further investigate the use of novel metabolism-altering therapies along with immunotherapy in medulloblastoma.

DON is a naturally occurring antibiotic first isolated from Streptomyces species in the 1950s [13]. It has been used in early phase clinical trials, including a phase I clinical trial in pediatric patients, however, it has never been systematically tested against MYC-expressing malignancies [41]. DON is known to target multiple enzymes that use glutamine, including glutaminase (GLS), asparagine synthetase, and phosphoribosylformylglycinamidine synthase [1,42]. The well-established linkages between MYC and glutamine metabolism in general [8], and the increased mortality observed in pediatric medulloblastoma patients with evidence of highly active tumor glutamine metabolism, [43] led us to hypothesize that DON prodrugs would be active against MYC-driven medulloblastoma models. We found that DON prodrugs were active at concentrations nearly 10-fold lower (10 uM vs 900 uM) than that recently reported for DON against the non-MYC medulloblastoma cell lines UW228 and DAOY [32], suggesting that the presence of the MYC proto-oncogene renders cells highly sensitive to glutamine metabolic inhibition. Indeed, DON prodrug concentrations that are highly lethal to neural stem cells engineered to express MYC have no effect on similar cerebellar stem cells immortalized with SV40. The addition of MYC to neural stem cells is engineered to express MYC have no effect on similar cerebellar stem cells.

We here demonstrate that oral JHU-083 provides robust delivery of MYC to transgenic medulloblastoma mice from late-stage cerebral malaria. [32]

References


